EP162-EN.01

OX40[Biotinylated]: OX40 Ligand Inhibitor Screening ELISA Kit

Pack Size: 96 tests

Catalog Number: EP-162

IMPORTANT: Please carefully read this manual before performing your experiment.

For Research Use Only. Not For Use In Diagnostic Or Therapeutic Procedures

INTENDED USE

This kit is designed for screening of inhibitors of binding between human OX40 and Human OX40 Ligand.

It is intended for research use only (RUO).

PRINCIPLE OF THE ASSAY

This inhibitor screening ELISA kit is designed to facilitate the identification and characterization of new OX40 pathway inhibitors. The assay takes advantage of our in house-developed binding of biotinylated human OX40 to immobilized human OX40 Ligand in a functional ELISA assay and employs a simple colorimetric ELISA platform. Briefly, we provide you with a human Biotinylated OX40 protein, a human OX40 Ligand protein, an anti-OX40 neutralizing antibody (as method verified Std.), and Streptavidin-HRP reagent. Your experiment will include 4 simple steps:

1) Coat the plate with human OX40 Ligand.

2) Add your molecule of interest to the tests.

3) Add biotinylated human OX40 to bind the coated human OX40 Ligand.

4) Add Streptavidin-HRP followed by TMB or other colorimetric HRP substrate.

Finally, the half maximal inhibitory concentration (IC50) of your compound to OX40: OX40 Ligand binding will be determined by comparing OD readings among different experimental groups.

MATERIALS PROVIDED

TABLE 1. MATERIALS PROVIDED (pls modify according to COA)

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Catalog	Components	Size (96 tests)	Format	Storage	
EP162-C01	High-bind Plate	1 plate	Solid	2-8℃	
EP162-C02	Human OX40 Ligand	10μg	Powder	2-8℃	-70℃ after reconstitution, avoid freeze-thaw cycles
EP162-C03	Biotinylated Human OX40	10μg	Powder	2-8℃	
EP162-C04	Anti-OX40 Neutralizing Antibody	10μg	Powder	2-8℃	
EP162-C05	Streptavidin-HRP	5μg	Powder	2-8℃, avoid light	
EP162-C06	Coating Buffer	12 mL	Liquid	2-8℃	
EP162-C07	20xWashing Buffer	50 mL	Liquid	2-8℃	
EP162-C08	Blocking Buffer	50 mL	Liquid	2-8℃	
EP162-C09	Substrate Solution	12 mL	Liquid	2-8℃, avoid light	
EP162-C10	Stop Solution	7 mL	Liquid	2-8℃	

REAGENTS/EQUIPMENT NEEDED BUT NOT SUPPLIED

Single or dual wavelength microplate reader with 450 nm/630nm filter;

Centrifuge;

37 ºC Incubator;

Single channel or multichannel pipettes with 10 μL, 200 μL and 1000 μL precision;

10 μL, 200 μL and 1000 μL pipette tips;

Test Tubes;

Graduated cylinder;

Deionized or distilled water for dilution;

STORAGE AND VALIDITY INSTRUCTIONS

Unopened kit should be stored at 2°C -8°C upon receiving. Find the expiration date on the outside packaging and do not use reagents past their expiration date.

The kit should be stored as TABLE 1 after the reconstitution of lyophilized materials. The shelf life is 30 days from

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the date of opening.

Note:

a. Do not use reagents past their expiration date.

b. Find the expiration date on the outside packaging.

REAGENT PREPARATION

1. Restore all reagents and samples to room temperature (20-25℃) before use.

2. Reconstitute the provided lyophilized materials to stock solutions with sterile deionized water as recommended in Tab.2, Solubilize for 15 to 30 minutes at room temperature with occasional gentle mixing. Avoid vigorous shaking or vortex. The reconstituted stock solutions should be stored at -70℃. **Avoid freeze-thaw cycles**.

Note: Streptavidin-HRP stock solution should be protected from light.

TABLE 2. RECONSTITUTION METHODS FOR 96 TESTS

Catalog	Components	Amount	Stock Solution Con.	Reconstitution Buffer and Vol.
EP162-C02	Human OX40 Ligand	10μg	100μg/mL	100μL, water
EP162-C03	Biotinylated Human OX40	10μg	100μg/mL	100μL, water
EP162-C04	Anti-OX40 Neutralizing Antibody	10μg	100μg/mL	100μL, water
EP162-C05	Streptavidin-HRP	5μg	50μg/mL	100μL, water

RECOMMENDED PROTOCOL

1. Working solution preparation

1.1 Preparation of 1×Washing Buffer:

Dilute 25 mL 20×Washing Buffer with ultrapure water/deionized water to 500mL.

1.2 Preparation of Dilution Buffer:

10 mL Blocking Buffer (EP162-C08) add 30 mL 1×Washing Buffer.

2. Coating

1)Dilute Human OX40 Ligand stock solution (100μg/mL) to 0.5μg/mL with Coating Buffer to make Human OX40

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Ligand working solution.

2)Add 100μL of Human OX40 Ligand working solution (0.5μg/mL) to each well and leave a couple of wells uncoated for No-Coating Control, seal the plate with microplate sealing film and incubate overnight (or 16 hours) at 4℃.

3. Washing

Remove the remaining solution by aspiration, add 300μL of 1×Washing Buffer to each well, gently tap the plate for 1 minute, remove any remaining 1×Washing Buffer by aspirating or decanting, invert the plate and blot it against paper towels. Repeat the washing step above for three times.

Note: For best results, the complete removal of the Human OX40 Ligand solution is essential. The use of a manifold dispenser or an auto-washer may be necessary.

4. Blocking

Add 300μL Blocking Buffer to each well, seal the plate with microplate sealing film and incubate at 37℃ for 1.5 hours.

5. Washing

Repeat step 3. At the same time, you can start to prepare your samples.

6. Add Samples

1)Make serial dilution of the samples as appropriate.

2)If you intend to use the provided Anti-OX40 Neutralizing Antibody as a reference (Std.), you may dilute the antibody as recommended in Figure 1.

3)Add 50μL of sample solution to each well according to our recommendation (Figure 2) or your own plate setup.

4)For No-Coating Control wells, please add 50μL Dilution Buffer.

7. Binding

1) Dilute Biotinylated Human OX40 stock solution (100μg/mL) to 0.08μg/mL with Dilution Buffer to make Biotinylated Human OX40 working solution.

2) For No-binding control wells, please add 50μL Dilution Buffer.

3) For all other wells, please add 50μL Biotinylated Human OX40 working solution to the wells and mix the samples by gently tapping the plate. Seal the plate with microplate sealing film and incubate at 37℃ for 1 hour.

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Note: The working solution should be prepared immediately before use and should not be stored.

FIG.1 PREPARATION OF 1:1 SERIAL DILUTIONS OF THE Anti-OX40 Neutralizing Antibody

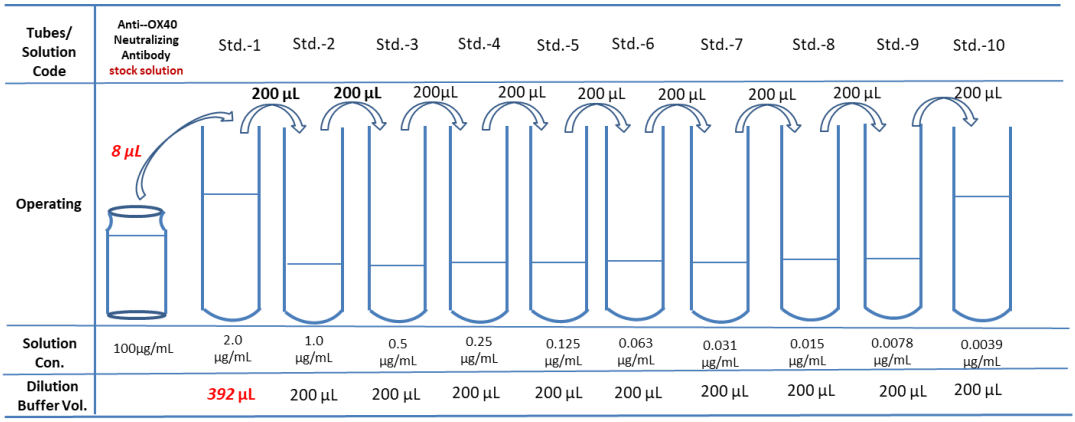
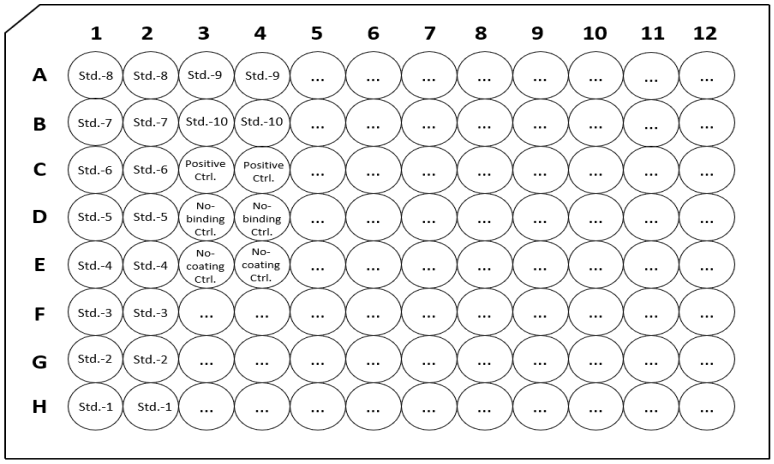


FIG.2 PLATE LAYOUT



8.Washing

Repeat step 3.

9.Add Streptavidin-HRP

1)Dilute Streptavidin-HRP stock solution (50µg/mL) to 0.1µg/mL with Dilution Buffer to make Streptavidin-HRP working solution.

2)For all wells, add 100µL Streptavidin-HRP working solution, seal the plate with microplate sealing film and incubate at 37℃ for 1 hour, avoid light.

10.Washing

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Repeat step 3.

11.Substrate Reaction

Add 100µL **Substrate Solution** to each well. Seal the plate with microplate sealing film and incubate at 37℃ for 20 minutes. Avoid light.

12.Termination

Add 50µL **Stop Solution** to each well, and gently shake the plate to allow thorough mixing.

Note: *the color in the wells should change from blue to yellow.*

13.Data Recording

Read the absorbance at 450 nm using UV/Vis microplate spectrophotometer.

Note*: Subtracting the value read at* OD_{450nm} *with* OD_{630nm} *can be used to reduce the background noise.*

SIMPLIFIED PROTOCOL

TABLE. 3 ASSAY PROTOCOL

Steps Code	Steps	Reagents & Instruments	Reaction Conditions	Samples	No-binding Ctrl.	No-coating Ctrl.	Positive Ctrl.
1	Working fluid preparation	N/A	N/A	N/A	N/A	N/A	N/A
2	Coating	Human OX40 Ligand Working Solution	4℃ for overnight	100µL	100µL	—	100µL
3	Washing	1XWash Buffer	Wash for 3 times	300µL	300µL	300µL	300µL
4	Blocking	Blocking Buffer	37℃ for 1.5 hours	300µL	300µL	300µL	300µL
5	Washing	1XWash Buffer	Wash for 3 times	300µL	300µL	300µL	300µL
6	Add Samples	Samples	—	50µL	—	—	—
		Dilution Buffer		—	50µL	50µL	50µL
7	Binding	Biotinylated Human OX40 Working Solution	Mix by gentle tapping, incubate at 37℃ for 1 hours	50µL	—	50µL	50µL
		Dilution Buffer		—	50µL	—	—
8	Washing	1XWash Buffer	Wash for 3 times	300µL	300µL	300µL	300µL

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9	Streptavidin-HRP	Streptavidin-HRP Working Solution	37℃ for 1 hours	100μL	100μL	100μL	100μL
10	Washing	1XWash Buffer	Wash for 3 times	300μL	300μL	300μL	300μL
11	Substrate Reaction	Substrate Solution	37℃ for 20 minutes	100μL	100μL	100μL	100μL
12	Termination	Stop Solution	Mix by gentle tapping	50μL	50μL	50μL	50μL
13	Data Recording	UV/Vis spectrophotometer	Measure absorbance at 450 nm, with the correction wavelength set at 630 nm				

Note for TAB. 3:

1) *Samples: Your samples of interest.*
2) *No-binding Ctrl.: Reaction without Biotinylated Human OX40 added. The absorbance should be around 0.05(< 0.1) at 450 nm.*
3) *No-coating Ctrl.: Reaction without Human OX40 Ligand coated on the wells. The absorbance should be around 0.05(< 0.1) at 450 nm.*
4) *Positive Ctrl.: Determined the max value in 450nm absorbance, when out of inhibitors.*
5) *It is recommended that all samples, controls and standards should be done in duplicates.*

PRECAUSIONS

1. This kit is for research use only and is not for use in diagnostic or therapeutic applications.
2. This kit should be used according to the provided instructions.
3. Do not mix reagents from different lots.
4. All reagents should be balanced to room temperature (20℃-25℃) before use.
5. This kit should be stored at 2℃-8℃.
6. Please prepare the working solution of each component according to the needs of the experiment. Except for 1x Washing Buffer, all prepared working solution is for one-time use and cannot be stored.

METHOD VERIFICATION

INHIBITION OF OX40[Biotinylated]: OX40 Ligand BINDING BY ANTI-OX40 NEUTRALIZING ANTIBODY

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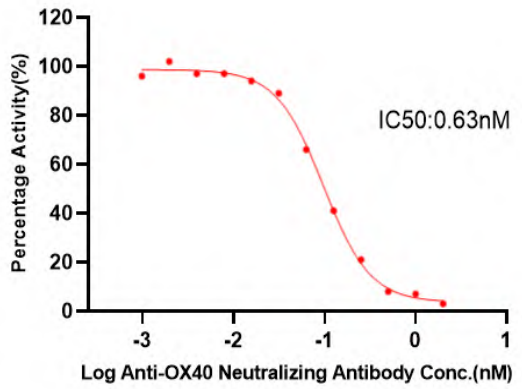
Serial dilutions of Anti-OX40 Neutralizing antibody (Catalog # EP162-C04) (1:1 serial dilution, from 2μg/mL to 0.001μg/mL) was added into OX40[Biotinylated]: OX40 Ligand binding reactions. The assay was performed according to the protocol described below. Background was subtracted from data points prior to log transformation and curve fitting.



Anti-OX40 Neutralizing Antibody Conc.(μg/ml)	Anti-OX40 Neutralizing Antibody Conc.(nM)	Mean Abs.(OD450)	Percentage Activity(%)
0	0.000	2.596	100%
0.0039	0.026	2.527	97%
0.0078	0.052	2.503	96%
0.0156	0.104	2.45	94%
0.0313	0.208	2.302	89%
0.0625	0.417	1.704	66%
0.125	0.833	1.057	41%
0.25	1.667	0.551	21%
0.5	3.333	0.217	8%
1	6.667	0.174	7%
2	13.333	0.089	3%
No Coating		0.055	
No Binding		0.064	

For each experiment, a standard curve needs to be set for each micro-plate, and the specific OD value may vary depending on different laboratories, testers, or equipments. The example data is for reference only.

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