

# **Influenza B/Victoria Lineage Viruses Hemagglutinin (HA) Specific ELISA Kit**

**Pack Size: 96 tests**

**Catalog Number: RAS-A206**

**IMPORTANT: Please carefully read this manual before performing your experiment.**

***For Research Use Only. Not For Use in Diagnostic or Therapeutic Procedures***

**INTENDED USE**

This kit is developed for specific quantitative detection of influenza B/Victoria lineage viruses HA protein in samples. This kit has been tested to specifically identify influenza B [Austria/1359417/2021 (B/Victoria lineage)] HA protein. It is intended for research use only (RUO).

**PRINCIPLE OF THE ASSAY**

Influenza, commonly known as 'the flu', is an infectious disease of birds and mammals caused by RNA viruses of the family Orthomyxoviridae, the influenza viruses. The virus is divided into three main types (Influenzavirus A, Influenzavirus B, and Influenzavirus C), which are distinguished by differences in two major internal proteins (hemagglutinin (HA) and neuraminidase (NA), which are the most important targets for the immune system. Hemagglutinin binds to the sialic acid-containing receptors on the surface of host cells during initial infection and at the end of an infectious cycle which makes it a great target for vaccine studies.

This assay kit is used to measure the levels of influenza B/Victoria lineage viruses HA protein by employing a standard sandwich-ELISA format. The microplate in the kit has been pre-coated with Anti-HA (Influenza B/Victoria Lineage) Antibody. First add the standard samples provided in the kit and your samples to the plate, incubate and wash the wells. Then add the HRP-Anti-HA (Influenza B/Victoria Lineage) Antibody to the plate, incubate and wash the wells. Lastly load the substrate into the wells and monitor color development in proportion with the amount of influenza B/Victoria lineage viruses HA protein present. The reaction is stopped by the addition of a stop solution and the intensity of the absorbance can be measured at 450 nm and 630 nm. The OD Value reflects the amount of influenza B/Victoria lineage viruses HA protein bound.

**MATERIALS PROVIDED**

**TABLE 1. MATERIALS PROVIDED**

Catalog	Components	Size (96 tests)	Format	Storage	
				Unopened	Opened
RAS206-C01	Pre-coated Anti-HA (Influenza B/Victoria Lineage) Antibody Microplate	1 plate	Solid	2-8°C	2-8°C
RAS206-C02	HA (Influenza B/Victoria Lineage) Standard	20 µg	Powder	2-8°C	-70°C

RAS206-C03	HRP-Anti-HA (Influenza B/Victoria Lineage) Antibody	20 µg	Powder	2-8°C, avoid light	-70°C, avoid light
RAS206-C04	10×Washing Buffer	50 mL	Liquid	2-8°C	2-8°C
RAS206-C05	2×Dilution Buffer	50 mL	Liquid	2-8°C	2-8°C
RAS206-C06	Substrate Solution	12 mL	Liquid	2-8°C, avoid light	2-8°C, avoid light
RAS206-C07	Stop Solution	7 mL	Liquid	2-8°C	2-8°C

### **REAGENTS/EQUIPMENT NEEDED BUT NOT SUPPLIED**

Single or dual wavelength microplate reader with 450 nm and 630 nm filter;

Centrifuge;

37°C Incubator;

10 µL, 200 µL and 1000 µL precision pipettes;

10 µL, 200 µL and 1000 µL pipette tips;

Multichannel pipettes;

Tubes;

Graduated cylinder to prepare Wash Solution;

Deionized or distilled water to dilute 10×Washing Buffer;

### **STORAGE**

1. Unopened kit should be stored at 2°C-8°C upon receiving.
2. Find the expiration date on the outside packaging and do not use reagents past their expiration date.
3. The opened kit should be stored per components table. The shelf life is 30 days from the date of opening.

### **REAGENT PREPARATION**

1. Bring all reagents and samples to room temperature (20 °C-25 °C) before use. If crystals have formed in buffer solution, place the sample in a 37 °C incubator until the crystals have completely dissolved and bring the solution back to room temperature before use.
2. Reconstitute the provided lyophilized materials to stock solutions with distilled, sterile water as recommended in

Table 2 and place the materials for 15 to 30 minutes at room temperature with occasional gentle mixing. Avoid vigorous shaking. The reconstituted stock solutions should be stored at -70°C. It is recommended not to freeze-thaw more than 1 time, the packing specification shall not be less than 5 µg.

**TABLE 2. RECONSTITUTION METHODS FOR 96 TESTS**

ID	Components	Size	Stock Solution Con.	Reconstitution Buffer and Vol.
RAS206-C02	HA (Influenza B/Victoria Lineage) Standard	20 µg	100 µg/mL	200 µL water
RAS206-C03	HRP-Anti-HA (Influenza B/Victoria Lineage) Antibody	20 µg	100 µg/mL	200 µL water

**RECOMMENDED SAMPLE PREPARATION**

**1. Working Fluid Preparation**

1.1 Preparation of 1×Washing Buffer:

Dilute 50 mL 10×Washing Buffer with ultrapure water/deionized water to 500 mL.

1.2 Preparation of 1×Dilution Buffer:

Dilute 50 mL 2×Dilution Buffer with 1×Washing Buffer to 100 mL.

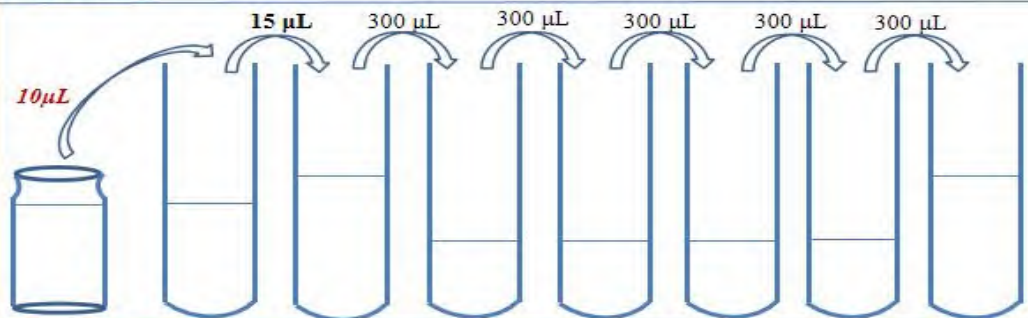
1.3 Preparation of HRP-Anti-HA (Influenza B/Victoria Lineage) Antibody working fluid:

Dilute HRP-Anti-HA (Influenza B/Victoria Lineage) Antibody to 1.0 µg/mL with Dilution Buffer. The prepared working fluid should avoid light. Please prepare it for one-time use only.

**2. Preparation of Standard Curve**

Make serial dilutions of the HA (Influenza B/Victoria Lineage) as a Standard curve with Dilution Buffer as recommended in Figure 1.

**FIGURE 1. PREPARATION OF 1:1 SERIAL DILUTIONS OF THE HA (Influenza B/Victoria Lineage)**

Tubes/ Solution Code	HA (Influenza B/Victoria Lineage) Standard stock solution	Std.-0	Std.-1	Std.-2	Std.-3	Std.-4	Std.-5	Std.-6
Operating								
Solution Con.	100µg/mL	2000 ng/mL	50 ng/mL	25 ng/mL	12.5 ng/mL	6.25 ng/mL	3.125 ng/mL	1.563 ng/mL
Dilution Buffer Vol.		490 µL	585 µL	300 µL	300 µL	300 µL	300 µL	300 µL

### 3. Add Samples

Add 100µL serially diluted **HA (Influenza B/Victoria Lineage)** Standard curve and samples to each well. For blank Control wells, please add 100µL 1×Dilution Buffer. Seal the plate with microplate sealing film and incubate at room temperature for 1.0 hour.

*Note: It is recommended to set double holes for samples and standard curves to be tested.*

### 4. Washing

Remove the remaining solution by aspiration, add 300 µL of 1×Washing Buffer to each well, soak for 30s, remove any remaining 1×Washing Buffer: by aspirating or decanting, invert the plate and blot it against paper towels. Repeat the wash step above for three times.

### 5. Add HRP-Anti-HA (Influenza B/Victoria Lineage) Antibody

For all wells, add 100 µL **HRP-Anti-HA (Influenza B/Victoria Lineage) Antibody (dilute to 1.0 µg/mL)** working solution. Seal the plate with microplate sealing film and incubate at room temperature for 1.0 hour.

### 6. Washing

Repeat step 4.

### 7. Substrate Reaction

Add 100  $\mu$ L **Substrate Solution** to each well. Seal the plate with microplate sealing film and incubate at room temperature for 20 min, avoid light.

## 8. Termination

Add 50  $\mu$ L **Stop Solution** to each well and tap the plate gently to allow thorough mixing.

*Note: the color in the wells should change from blue to yellow.*

## 9. Data Recording

Read the absorbance at 450 nm and 630 nm using UV/Vis microplate spectrophotometer within 5 minutes.

*Note: To reduce the background noise, subtract the value read at  $OD_{450\text{ nm}}$  with the value read at  $OD_{630\text{ nm}}$ .*

## CALCULATION OF RESULTS

1. Normal range of Standard curve:  $R^2 \geq 0.9900$ , detection range: 1.563-50 ng/mL.
2. If the OD value of the sample to be tested is higher than the highest standard, the sample shall be diluted with dilution buffer and assay repeated.
3. To calibrate absorbance value obtained by the standard curve, the OD value of the sample to be measured is subtracted from the OD value of the blank control. The standard curve is plotted with the standard concentration as x-axis and the calibrated absorbance value as y-axis. Four parameters logistic are used to draw the standard curve and calculate the sample concentration.

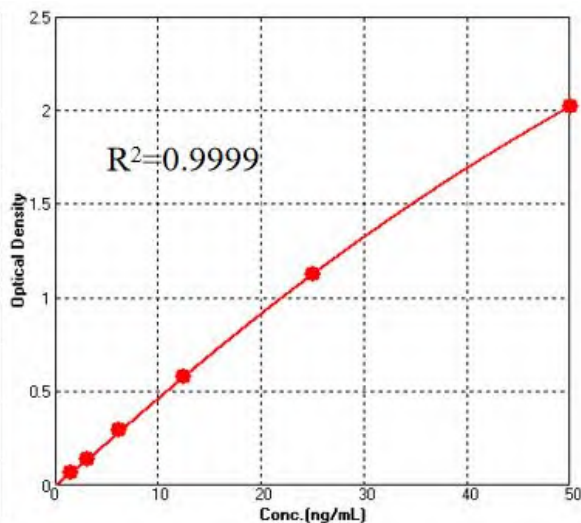
## PRECAUTIONS

1. This kit is for research use only and is not for use in diagnostic or therapeutic procedures.
2. The kit should be used according to the instructions.
3. Do not mix reagents from different lots.
4. Bring all reagents and samples to room temperature (20°C-25°C) before use. If crystals have formed in buffer solution, warm to room temperature until the crystals have completely dissolved.
5. The kit should be stored at 2°C to 8°C.

## TYPICAL DATA

The following data is for reference only. The sample concentration was calculated based on the results of the standard curve.

Standard (ng/mL)	O.D.-1	O.D.-2	Average	Corrected
50	2.118	2.030	2.074	2.024
25	1.150	1.208	1.179	1.129
12.5	0.611	0.651	0.631	0.581
6.25	0.331	0.352	0.342	0.292
3.125	0.181	0.190	0.186	0.136
1.563	0.120	0.122	0.121	0.071
0	0.050	0.050	0.050	/



## PRECISION

### 1. Intra-assay Precision

Three samples of known concentration were tested ten times on one plate to assess intra-assay precision.

### 2. Inter-assay Precision

Three samples of known concentration were tested in three separate assays to assess inter-assay precision.

Sample	Intra-assay Precision			Inter-assay Precision		
	1	2	3	1	2	3
n	10	10	10	3	3	3
Mean (ng/mL)	37.204	18.471	3.719	36.946	18.566	3.660
SD	1.255	0.820	0.183	2.460	0.256	0.051
CV (%)	3.4	4.4	4.9	6.7	1.4	1.4

*Note: The example data is for reference only.*

## RECOVERY

Three samples with different concentrations were tested to calculate the recovery rate.

Sample(n=5)	Average Recovery %	Range %
High	96.2	88.1-103.7
Middle	94.1	85.3-100.3

Low	94.2	81.8-113.1
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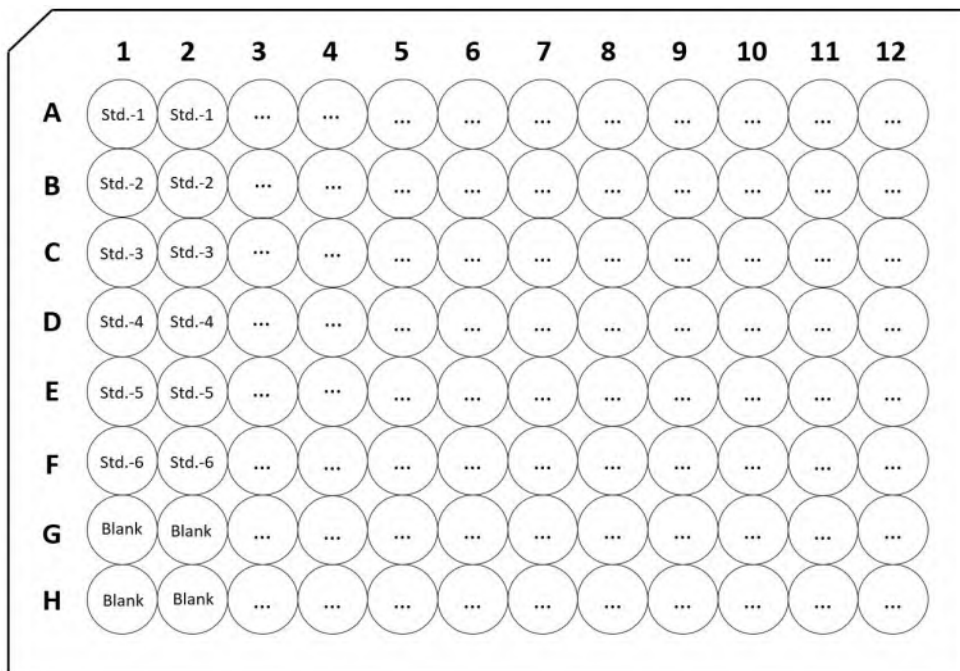
**LINEARITY**

To assess the linearity of the assay, samples spiked with high concentrations were serially diluted with calibrator diluent to produce samples with values within the dynamic range of the assay.

		Cell culture medium (DMEM)	Cell culture medium (1640)
1:2	Average Recovery (%)	95.6	93.1
	Range (%)	89.7-101.3	91.1-98.5
1:4	Average Recovery (%)	95.2	89.6
	Range (%)	89.7-103.9	85.0-97.1
1:8	Average Recovery (%)	92.0	92.0
	Range (%)	87.9-97.7	88.9-97.9
1:16	Average Recovery (%)	94.0	92.0
	Range (%)	87.3-98.1	85.0-97.2

*Note: The example data is for reference only.*

**PLATE LAYOUT**





*Note: Blank is a Blank Dilution Buffer hole.*

**TROUBLESHOOTING GUIDE**

<b>Problem</b>	<b>Cause</b>	<b>Solution</b>
<b>Poor standard curve</b>	* Inaccurate pipetting	* Check pipettes
<b>Large CV</b>	* Inaccurate pipetting * Air bubbles in wells	* Check pipettes * Remove bubbles in wells
<b>High background</b>	* Plate is insufficiently washed * Contaminated wash buffer	* Review the manual for proper wash. * Make fresh wash buffer
<b>Very low readings across the plate</b>	* Incorrect wavelengths * Insufficient development time	* Check filters/reader * Increase development time
<b>Samples are reading too high, but standard curve looks fine</b>	* Samples contain cytokine levels above assay range	* Dilute samples and run again
<b>Drift</b>	* Interrupted assay set-up * Reagents not at room temperature	* Assay set-up should be continuous - have all standards and samples prepared appropriately before commencement of the assay * Ensure that all reagents are at room temperature before pipetting into the wells unless otherwise instructed in the antibody inserts